ϵ Singclean® **COVID-19 Antigen Test Kit**

(Colloidal Gold)

COVID-19 Antigen Test kit (Golioidal Gold) is a solid phase immunochromatographic assay for the rapid, qualitative detection of antigen to 2019 Novel Coronavirus in human saliva. This test provides only a preliminary test result. Therefore, any reactive specimen with the COVID-19 Antigen Test kit (Colloidal Gold) must be confirmed with alternative testing method (s) and clinical findings.

PACK FORMATS 1 test/box, 20 tests/box, 50 tests/box, 100 tests/box INTRODUCTION
The novel coronaviruses.

IN ROUDCION. The novel coronaviruses belong to the β genus. COVID-19 is an acute respiratory infectious disease. People are generally susceptible. Currently, the patients infected by the novel coronavirus are the main source of infection; asymptomatic only infected people can also be an infectious source. Based on the current epidemiological investigation, the incubation period is 1 to 14 days, mostly 3 to 7 days. The main manifestations include respiratory of the country of the

Services (Seption 1. the 'incubation period is 1 to 14 days, mostly 3 to 7 days. The main manifestations incuise incurse inclique and dry cough. Nasal congestion, runny nose, sore throat, myalgia and diarrhea are found in a few cases PRINCIPLE
The COVID-19 Antigen Test kit (Colloidal Gold) is a colloidal gold immunochromatographic assay. The test uses COVID-19 antibody (test tine 7) and goat anti-mouse polyclonal antibody (control line C) immobilised on a introcellulose strip. The burguindy colored conjugate pad contains colloidal gold conjugated to another COVID-19 antibody conjugated with colloid gold. When the processed buffer containing the sample is added to the sample elik. (CVID-19 will combine with the COVID-19 antibody complex magnets through introcellulose membrane by capillary action. When the complex meets the line of the event of the complex may be sent to the complex meets the line of the test result. Absence of a colored band in the test means negative test result. The test kit contains a quality control line (control line C), which should show a burgundy color band of goat anti-mouse polyclonal antibody combined with the colloidal gold conjugate in the gold label pad, regardless of the color on any other test line.

MATERIALS SUPPLED

Sealed pouches each containing a test cassette, a desiccant Sampling cotton swabs (as saliva swab)

Antigen extraction tube

Antigen extraction tube
Paper workbench (The small one-test-box can be used as a workbench)

MATERIALS REQUIRED BUT NOT PROVIDED

STORAGE AND STABILITY
The kit can be stored The kit can be stored at room temperature or refrigerated (4-30°C). The test device is stable through the expiration date printed on the sealed pouch. The test device must remain in the sealed pouch until use.

DO NOT FREEZE.

After opening the expiration date.

After opening the sealed pouch, use the test as soon as possible within 60 minutes.

WARNINGS AND PRECAUTIONS

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WARNINGS AND PRECAUTIONS

1. For professional In Vitro diagnostic use only. Do not use after expiration date.

2. These instructions must be strictly followed by a trained healthcare professional to achieve accurate results. All users have to read the instruction prior to performing a test.

3. Do not use it if the tube/pouch is damaged or broken.

4. Wear protective gloves while handling specimens and wash hands thoroughly afterwards.

5. Avoid splashing or aerosol formation of specimen and buffer.

6. Clear up splits thoroughly using an appropriate disinfectant.

7. Decontaminate and dispose of all specimens, reaction kits and potentially contaminated materials (such as according to applicable local regulations.

8. Do not mix reagent of different lots or those for other products.

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10. Do not store the test kit in direct sunlight.

11. To avoid contamination, do not touch the head of provided swab when opening the swab pouch.

12. The provided swabs in the package should be used only for saliva specimen collection.

13. To avoid cross-contamination, do not rouse the swabs for specimen collection.

14. Do not glittle the collected swab with any solution except for the provided extraction buffer.

15. Test is for single use only. Do not reuse the swabs for specimen collection.

16. Do not perform the test in a room with strong air flow, such as electric fan or strong air-conditioning.

17. Test is for single use only. Do not reuse under any circumstances.

18. Do not perform the test in a room with strong air flow, such as electric fan or strong air-conditioning.

18. Test is for single use only. Do not reuse under any circumstances.

18. Do not determine the memberature prior to testing.

28. Testing should be performed immediately after specimen collection.

29. Briting specimens to room temperature prior to testing.

21. Testing from the stricum temperatur

b. Avoid chewing or swallowing during the sampling period. The saliva sample should be tested immediately after sampling. TEST PROCEDURE
Allow test cassetle, Antigen extraction buffer, specimen and controls to equilibrate to room temperature (16:50°C) prior to testing.

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The saliva is a second residue in the mouth before sampling, if the patient just has eaten, gargle or tooth brushing is required. Remove the swab package, keep the swab on the tongue until the tip being fully soaking by saliva (at least two minutes).

2. Place the antigen extraction tube on the workbench. Place the antigen extraction buffer bottle vertically downward, squeeze the bottle to make the buffer drip freely into the antigen extraction tube without touching the edge of the tube, and add all of the antigen extraction to buffer on the sample of the swab specimen into the antigen extraction to tube pre-added with the antigen extraction buffer, and rotate the swab about 10 times withey pressing the swab head against the tube wall to release the antigen in the swab, then let It stand for about 1 minute.

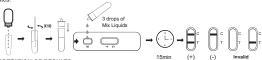
5. Install the dripper on the antigen extraction tube and cap it tightly, and let it stand for about 1 minute.

6. Remove the test cassette from the sealed foil pouch and use it as son as possible.

7. Place the test device on a clean and level surface.

8. Transfer 3 drops (about 1001) of mix Liquids to the sample well of the test card (or use a pipette to add 100ul), and start the timer.

9. Walt for the test result of the reagent. The result should be read in 15 minutes. Do not interpret the result after 20 minutes.



INTERPRETATION OF RESULTS

NEGATIVE:
If only the C band is present, the absence of any burgundy color in the T band indicates that no COVID-19 antigen is detected in the specimen. The result is negative.

POSITIVE:
In addition to the presence of C band, if T band is developed, the test indicates for the presence of COVID-19 antigen in the specimen. The result is COVID-19 positive.

INVALID:
Control line falls to appear, Insufficient specimen volume or incorrect procedural techniques are the most likely reasons for control line failure. Review the procedure and repeat the test with a new test cassette. If the problem persists, discontinue using the test kit immediately and contact your local distributor.

LIMITATIONS

1. Use fresh samples whenever nossible.

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1. Use fresh samples whenever possible.
2. Optimal assay performance requires strictly adherence to the assay procedure described in Instruction for use. Deviations may lead to aberrant results.
3. A negative result for an individual subject indicates absence of detectable COVID-19 antigen. However, a negative test result does not preclude the possibility of exposure to or infection with COVID-19.
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4. angulative test result does not preclude the covidence of the preclude of the patient of a single test, but should only be made by the priction after all clinical and aboratory findings have been evaluated.

1. Clinical Serviciality Secretificity and Accuracy.

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1. Clinical Sensitivity, Specificity and Accuracy Atotal of 397 samples were tested in this study. The results of test reagent and control reagent both were 230 negative specimens and 167 positive specimens. The data were collected in 2021.03.02–2021.03.11, and the sensitivity and specificity calculated were valid in this study.

Table 1: COVID-19 Artigen Test Rift vs PCR

Method		PCR results			
		Positive	Negative	Total	
COVID-19 Antigen Test kit	Positive	167	1	168	
	Negative	0	229	229	
Total		167	230	397	
Sensitivity		>99.99%	95% confidence interval	97.75%~100%	
Specificity		99.57 %	95% confidence interval	97.58%~99.92%	
Accuracy		99.75 %	95% confidence interval	98.59%~99.96%	

2.Limit of Detection (LOD)

LOD studies determine the lowest detectable concentration of SARS-CoV-2 at which approximately 95% of all (true positive) replicates test positive. Heat inactivated SARS-CoV-2 virus, with a stock concentration of 7.8 x 10′ TCID_mL, was spiked into negative specimen and serially diluted. Each dilution was ran in triplicate on the COVID-19 Test kit is 9.975 x 10² TCID_s/mL (Table 2). Table 2: Limit of Detection of the COVID-19 Test kit is 9.975 x 10² TCID_s/mL (Table 2).

Concentration	Concentration	Concentration
9.975 x 10 ² TCID ₅₀ /mL	20/20	100%

3.High Dose Hook Effect
No high dose hook effect was observed when testing up to a concentration of 7.8 x 10st TCID_{so}/mL of heat
inactivated SARS-Co-V2 virus.
4.Cross Reactivity
Cross reactivity with the following organisms has been studied. Samples positive for the following organisms
were found negative when tested with the COVID-19 Test Kit.
Table 3: Cross Reactivity City Results

Table 5: Cross Reactivity Stu	IV Results		
Pathogens	Concentration	Influenza A H5N1 virus	1.95 x 10 ⁶ TCID ₅₀ /mL
Human Coronavirus 229E	1 x 10° PFU /mL	Influenza B Yamagata	1.3 x 10 ⁶ TC I D ₅₀ /mL
Human Coronavirus OC43	1 x 10° PFU /mL	Influenza B Victoria	2.6 x 10 ⁶ TCID ₅₀ /mL
Human Coronavirus HKU1	1 x 10 ⁶ PFU/mL	Haemophilus influenzae	3.8 x 10 ⁶ PFU/mL
Human Coronavirus NL63	1 x 10 ^s PFU/mL	Rhinovirus (type 2)	1 x 10 ⁵ PFU/mL
Adenovirus (type 5)	1.8 x 10 ⁵ PFU/mL	Rhinovirus (type 14)	3.8 x 10 ⁵ PFU/mL
Adenovirus (type 7)	3.2 x 105 TCID _{so} /mL	Rhinovirus (type 16)	5.5 x 10 ⁶ PFU/mL
Adenovirus (type 18)	1.6 x 105 TCID _{sc} /mL	Respiratory syncytial virus(type A-2)	2.8 x 10 ⁵ PFU /mL
Human metapneumovirus (hMPV)	1.5 x 105 PFU/mL	Streptococcus pneumoniae	2.3 x 10 ⁵ PFU/mL
Parainfluenza virus (type 1)	1.8 x 105 TCID ₅₀ /mL	Streptococcus thermophilus	3.8 x 10 ⁵ PFU/mL
Influenza A H1N1 virus	2.1 x 105 TCID ₅₀ /mL	Mycoplasma pneumoniae	4.5 x 10° PFU/mL
Influenza A H3N2 virus	1.8 x 105 TCID _{sc} /mL	Chlamydia pneumoniae	6,3 x 10 ⁵ PFU/mL

The following substances, naturally present in respiratory specimens or that may be artificially introduced into the nasopharynx, were evaluated COVID-19 Test kit at the concentrations listed below and were found not to affect Table 4: Interfering Substance Study Results

Substance	Concentration	Ibuprofen	200μg/mL
Hemoglobin	2mg/mL	Morpholine Hydrochloride	200μg/mL
Throat lozenges	10mg/mL	Cephalexin	3μg/mL
Human Anti-mouse Antibody (HAMA)	5mg/L	Kanamycin	3µg/mL
Biotin	10mg/mL	tetracycline	3µg/mL
Mouthwash	500µl/mL	Chloramphenicol	3µg/mL
Gentamicin	3µg/mL	Erythromycin	3µg/mL
Sodium Cromdyn	120µg/mL	Vancomycin	3µg/mL
Oxymetazoline Hydrochloride	60µg/mL	Nalidixic acid	3µg/mL
Phenylephrine Hydrochloride	200µg/mL	Hydrocortisone	3µg/mL
N-Acetyl Paraaminophenol	200μg/mL	Human insu l in	3µg/mL
Aspirin	30µg/mL	Beta-propiolactone	30µg/mL

6. Microbial Interference

Microbial Interference with the detection of COVID-19 Test
Kill evaluate whether observation in clinical samples interfere with the detection of COVID-19 Test
Kill evaluate whether of false negative results. Each pathogenic microorganism was tested in triplicate in the
presence of heat inactivated SARS-Cov2 vins (9.975x10* TCID_{ss}/mL). No cross reactivity or interference was
seen with the microorganisms listed in the table below.

Table 5: Microbial Interference	Study Results		
Pathogens	Concentration	Influenza A H5N1 virus	1.95 x 10 ⁵ TCID ₅₀ /mL
Human Coronavirus 229E	1 x 10 ⁵ PFU /mL	Influenza B Yamagata	1.3 x 10 ⁵ TCID _{sc} /mL
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Human metapneumovirus (hMPV)	1.5 x 10 ⁵ PFU/mL	Streptococcus pneumoniae	2.3 x 10 ⁵ PFU/mL
Parainfluenza virus (type 1)	1.8 x 10 ⁵ TCID ₅₀ /mL	Streptococcus thermophilus	3.8 x 10 ⁵ PFU/mL
Influenza A H1N1 virus	2.1 x 10 ^s TCID _{ss} /mL	Mycoplasma pneumoniae	4.5 x 10° PFU/mL
Influenza A H3N2 virus	1.8 x 10 ^s TCID _{so} /mL	Chlamydia pneumoniae	6.3 x 10 ^s PFU/mL

REFERRENCE

1. Weiss SR, Leibowitz JL. Coronavirus pathogenesis. Adv Virus Res 2011; 81: 85-164.

2. Masters PS, Perham S. Coronavirdida: In: Knipe DM, Howley PM, eds. Fields virology. 6th ed. Lippincott Williams & Wilkins, 2013; 825-83.

3. Su S., Wong G, Shi NY, et al. Epidemiology, genetic recombination, and pathogenesis of coronaviruses. Trends Microbiol 2016; 24: 490-902.

Cui J, Li F, Shi ZL. Origin and evolution of pathogenic coronaviruses. Nat Rev Microbiol 2019; 17: 181-192.
 SYMBOLS USED ON PACKAGING NOTICE

EC REP	Authorized Representative	1	Store between 4-30°C	IVD	For in vitro diagnostic use only	[]i	Consult instructions for use
(2)	Do not reuse	LOT	Lot Number	(60)	Don't use if package is damaged	Ω	Use by



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